ASSESMENT OF THE THERAPUTIC EFECT OF LATE AZITHROMYCIN TREATMENT ON CHRONIC MURINE TOXOPLASMOSIS

By

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Abstract

Toxoplasmosis is a widespread, neglected disease with significant morbidity and mortality. The current study aimed to investigate the potential effectiveness of azithromycin against the cystogenic Me 49 nonvirulent strain of *Toxoplasma gondii* after the 6th week post-infection (PI) in chronically infected mice. Fifty laboratory-bred female Swiss albino mice were divided into five groups: G1: Non-Infected, G2: Infected non-treated, G3: Infected and treated by azithromycin 200 mg/kg after the 6th week PI for 3 days, G4: Infected and treated by azithromycin 200 mg/kg after the 6th week PI for 10 days and G5: Infected and treated by spiramycin 200 mg/kg after the 6th weeks, brain cysts number, size and histopathological changes were evaluated after 2 months.

The results showed that treating mice after the 6th week post-infection with azithromycin for 10 days reduced number and size of brain cysts by 64% & 30%, respectively, but without significant difference as compared with spiramycin-treated ones. Azithromycin treatment for 3 days achieved a greater reduction (34%) in brain cyst size than the group treated with azithromycin for 10 days. But, it caused the least reduction in brain cyst number (10%).Azithromycin treatment for 10 days improved the histopathological changes in cerebral toxoplasmosis.

Keywords: Mice; Azithromycin; Spiramycin; Toxoplasma gondii; Treatment

Introduction

Toxoplasmosis is an infection with an intracellular parasite called Toxoplasma gondii that affects about one-third of the population, exists in nature as oocysts, bradyzoites (in latent tissue cysts) and replicating tachyzoites, which represents the hallmark of active disease (Duffy et al, 2019). Infection occ urs through several routes, including ingestion of contaminated water or food (as main route), contact with cat litter contaminated with oocysts, blood transfusion, organ/tissue transplantation (Maleki et al, 2021) and congenital infection from infected mother to her fetus via placenta (Morsy et al, 2022). Toxoplasmosis disrupts fetal brain development and severely affects the host's brain, has been linked to many behavioral and neurological disorders (Abo-Al-Ela, 2020). Also, t toxoplasmosis is a probable risk factor for neurodevelopmental disorders in children. This highlights the importance of toxoplasmosis consideration by pediatricians for investigating and early management of congenital and neurodevelopmental disorders (El-Beshbishi *et al*, 2020). The unique pathogenesis of *T. gondii* also presents challenges for drug therapy, in contrast to other apicomplexans, penetrates the blood-brain barrier and creates a bradyzoite infection that was drug resistant (Alday and Doggett, 2017).

Treatment guidelines for toxoplasmic encephalitis recommend either pyrimethamine and sulfadiazine or pyrimethamine and clindamycin; trimethoprim/sulfamethoxazoleole is additionally known to possess comparable potency (Hernandez *et al*, 2017). Pyrimethamine and sulfadiazine, however, were linked to rare severe responses that may be fatal, such as agranulocytosis, Stevens-Johnson syndrome, toxic epidermal necrolysis or hepatic necrosis (Harr and French, 2010). The development of anti-*T. gondii* treatment with activity against cysts during the latent stage of infection is greatly required to diminish the cyst load within the host and may eliminate the parasite from the host to achieve sterile immunity (Dunay et al, 2018). The primary factor influencing the ongoing interest in azithromycin is its unique pharmacokinetic characteristics. In short, azithromycin is considerably stable at low pH, avaried antimicrobial spectrum, a long half-life, excellent tissue penetration and extensive tissue distribution and great drug concentrations within cells (including phagocytes) (McMullan and Mostaghim, 2015). Regarding the mechanism of action of azithromycin, it was found that in bacteria, azithromycin binds to the RNA of the 50S ribosomal subunit and acts synergistically to prevent protein synthesis (Beckers et al, 1995). Azithromycin also destroys tissue cysts in addition to its anti-replication effect on T. gondii tachyzoites; it has good intracellular penetration and can affect intracellular tachyzoites (Bosch-Driessen et al, 2002). Azithromycin has anti-inflammatoory and immunom odulatory activities (Kanoh and Rubin, 2010). Azithromycin exhibits bacteriostatic capabilities against a wide range of gram-positive, gram-negative bacteria, and even atypical bacteria (McMullan and Mostaghim, 2015). Moreover, it has anti-parasitic activity against cryptosporidiosis (Kadappu et al, 2002), Plasmodium species (Srivastava et al, 2012) and Trypanosoma brucei brucei (Molefe et al, 2019), as well as was employed in Egypt as an antiviral drug for COVID-19 (Kamel et al, 2022).

The current study aimed to investigate the possible therapeutic effect of azithromycin against the cystogenic Me-49 non-virulent strain of *Toxoplasma gondii* in chronically infected mice.

Materials and Methods

Parasite: *T. gondii* Me 49 non-virulent strain was kindly provided by Zoonosis Department, National Research Center, and regularly maintained by sub passage in Swiss Alb ino mice with 0.1 ml of brain homogenate at 1×10^2 tissue cysts/ml every 8 weeks to develop chronic toxoplasmosis. Brains were ground with sterile pestle in a clean mortar and diluted 1×10^2 cysts/ml brain cyst suspension. Drugs: Two drugs were used.1-Spiramycin[®] (Spirex) and 3 MIU (704mg) was purchased from Medical Union Pharmaceuticals, Egypt. Tablets were crushed and dissolved in distilled water for oral suspension at a dose of 200 mg/kg/day (Grujić*et al*, 2005).

2- Azithromycin (Zithromax, Pfizer Egypt) was prepared daily as a liquid suspension. After sonication, the homogenized suspensions were given orally to mice by stomach tube at a dose of 200mg/kg day. Oral dose was 0.1 ml/mouse.

Experimental infection: Mice were inoculated orally with 0.1ml of brain cysts suspensioncontained 10 cysts for every mouse.

Experimental Design: Fifty clean laboratory-bred female Swiss Albino mice, 10 weeks old and weighed 20-25gm were classified into five groups of 10 mice in each: G1: Non-infected (control negative), G2: Infected non-treated (control positive), G3: Infected and treated with azithromycin 200 mg/kg after the 6th week post-infection (PI) for 3 days, G4: Infected and treated with azithromycin 200 mg/kg after 6thweek PI for 10 days, G5: Infected and treated with spiramycin 200 mg/kg after 6th week PI for 2 weeks.

Assessment of anti-*Toxoplasma* action: After 8 weeks, all mice were sacrificed and their brains were removed. Each brain was separated into two sections, one for counting the cysts and the other was fixed in 10% formalin for the histopathological studies.

Brains were harvested, rinsed in sterile normal saline solution, weighed, and 1ml of sterile saline was added, followed by homogenization (Omni TH-220) for 5 minutes.

Histopathological examination for cysts: Brain specimens were fixed in 10% neutral buffered formalin, and processed for paraffin sections of 5µm thickness, stained with haematoxylin and eosin (H & E) and microscopically examined (Gad *et al*, 2022). Besides, homogenate brain (0.1ml) smear was spread on a clean slide, fixed in methanol, air dried and stained in Giemsa stain (Merck, Germany) for 30-45 minutes, rinsed with water, dried, and microscopically examined for cyst counts. The following equation was used: Mean cyst number = cyst count in $100\mu \times 10 \times 2$. Ocular and stage micrometers were used to count brain cyst and measure size.

Ethical consideration: The Research Ethics Committee, Benha Faculty of Medicine, revised and approved the protocol (Approval No. Ms14-11-2018), and the Albino mice experimented with was done following the internationally standards of Helsinki declaration (2008).

Statistical analysis: Data were coded, tabulated and analyzed using ANOVA procedure and the post hoc test (Feldman *et al*, 2003).

Results

Both azithromycin and spiramycin significantly reduced the number of Toxoplasma brain cysts when compared with the control infected non treated group except the group treated with azithromycin for only 3 days. Mice group treated with azithromycin for 10 days caused more reduction in cyst number than those treated for 3 days (P<0.05). The number of Toxoplasma brain cysts achieved by the group treated with azithromycin for 3 days was much lower than those treated with spiramycin for 2 weeks (P<0.05). There was no difference in the outcome of treatment with the group of azithromycin treated for 10 days when compared with the group of spiramycin (P>0.05).

All azithromycin and spiramycin-treated mice exhibited significant reduction in the size of brain cysts compared with the control positive group. Azithromycin treatment for 10 days resulted in a smaller reduction in the size of brain cysts than spiramycin treatment (P<0.05). When mice treated with azithromycin for three days was compared to those treated with spiramycin, no significant redu-

ction in brain cysts size (P>0.05). There was no statistically significant reduction in the size of brain cysts when the two azithromycin groups were compared (P>0.05).

Histopathological results: Brains of control negative mice showed normal histological structures. But, brain sections from positive control showed severe histopathological changes. Examined sections showed noticeable severe necrosis of neurons and neuronophagia with the presence of neurofibrillary tangles and numerous large Toxoplasma cysts distributed throughout the brain parenchyma. Noticeable vasculitis and perivasculitis as well as numerous focal necrosis associated with glial cell infiltration and severe perivascular cuffing with mononuclear inflammatory cells were also detected. Microscopically, examined brain sections from mice treated with azithromycin for 3 days after 6th week showed moderate to severe necrosis of neurons, neuronophagia, neurofibrillary tangles, marked perivascular cuffing with mononuclear cells, moderate meningitis (inflammatory infiltrate in meninges) associated with Toxoplasma cysts and several different Toxoplasma developmental stages in cerebral cortex. Also, examination of brain sections from azithromycin-treated mice for 10 days showed several alterations such as necrosis of neurons with neurofibrillary tangles appearance and neuronophagia as well as glial nodules (focal gliosis). Few Toxoplasma cysts and intravascular bradyzoites were also detected, without meningitisbut mild perivascular cuffing with mononuclear cells. Brains of mice treated with spiramycin for 2 weeks PI showed focal infiltration with inflammatory cells and marked necrosis of some neurons. Details were given in table (1) and figure (1).

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Group	Drugs	Dose/day	Treatment	Dose	No of T. gondii cyst			size of T. gondii cyst		
		(mg/kg)	duration (d)	(mg/kg)	M±SD	Reduct %	95% CI	M±SD	Reduct%	95% CI
G2:	N/D	N/D	N/D	N/D	526±40	N/D	N/D	24±4	N/D	20-27
G3:	Azithr.	200	3	600	482±69	10	433-531	16±2	34	14-16
G4:	Azithr.	200	10	2000	192±30	64	170-214	17±2	30	16-19
G5:	Spira.	200	14	2800	148 ± 48	72	113-183	14±2	42	13-15

Table 1: Effect of Azithromycin on number and size of T. gondii brain cyst in chronic infected mice

Azithro. = azithromycin Spira.= Spiramycin Reduct + Reduction

Discussion

In the present study, treatment regimen 200 mg per kg per day azithromycin for 10 days showed an obvious therapeutic effect in treating the chronic toxoplasmosis infection among the studied groups, as the brain cyst number reduction rate was 64%. There was no significant difference with spiramycintreated group (72% reduction rate). Conversely, the group treated with 200mg/kg/day azithromycinforonly3days had the lowest rate of brain cyst reduction among all groups (10%). Araujo et al. (1988) evaluated azithromycin treatment with a dose of 200mg/kg/ day for 10 days in mice infected with strain C56 of T. gondii, treatment entirely protected mice from death induced by T. gondii intraperitoneal infection. Also, Araujo et al. (1991) reported that in toxoplasmosis, the tissue concentrations of azithromycin in the brain were ten-fold greater than the concentrations in the serum after treatment with 200mg/kg/ day for 10 days (i.e., a dose regimen comparable to the present study used). Azithromycin monotherapy at a dose of 500mg/day was shown to be effective and well-tolerated for the treatment of active, non-vision-threatening toxoplasmic retinochoroiditis, also, duration of treatment was clinically longer for the azithromycin group (Balaskas et al, 2012). This agreed with the present data that the concentrations in infected mouse brains were approximately twofold greater than those in the brains of non-infected mice.

The present data also agreed with Lescano *et al.* (2004) investigated azithromycin treatment at a dose of 100mg/kg/day orally 20 days after infection and continued for 120 days. Number of brain cysts in mice infected intraperitoneally with a cystogenic *T. gondii* strain was reduced by 45% as compared to positive control group. Besides, Montaya and Remington (2008), who reported that azithromycin, especially at 200mg/ kg/day, was effective in both prophylaxis and treatment of toxoplasmosis due to its ability to reach significant concentrations in the CNS. Previously, mice given azithromycin and IFN

survived at a rate of <40%; but, azithromycin alone protected less than 10% of mice, and IFN alone didn't protect against a lethal dosage of *T. gondii* (Araujo and Remington, 1991). Besides, Dumas *et al.* (1994) treated mice with azithromycin at a dose of 100mg/ kg /day for 100 days beginning 4 weeks after infection didn't find significant change in the brain cysts number compared to the positive control group. The reduced severity of the retinal lesions and improvement vision by azithromycin was found to be a satisfactory as an alternative regimen to trimethoprim/sulfam-ethoxazole (Lashay *et al*, 2016).

The current study showed that the Albino mice treated with spiramycin for 2 weeks had the greatest reduction in the size of *Toxoplasma* brain cysts (42%), followed by mice treated with azithromycin for 3 days (34%), and the group treated with azithromycin for 10 days had the least reduction. However, many macrolides inhibited *T. gondii* intracellular growth; azithromycin was the only macrolide that inhibited intracellular tachyzoite multiplication (Chamberland *et al*, 1991).

Dumas et al. (1994) and Derouin (1995) found that azithromycin is also effective against intracystic bradyzoites in-vitro, but long-term administration of the drug to chronically infected mice failed to reduce the average number of cysts in the brain, but reduced the inflammatory response was noticed in the brains of infected and treated animals. This could be due to azithromycin's antireplication effect on T. gondii tachyzoites, which even destroyed tissue cysts. Azithromycin also possesses good intracellular penetration and may directly affect intracellular tachyzoites (Bosch-Driessen et al, 2002). Besides, Degerli et al. (2003) reported that the azithromycin acted against the T. gondii tachyzoites and cysts as well as bradyzoites

The fluctuation in size of *Toxoplasma* cysts discovered in the current study revealed that chronic infection resulted in the continued creation of new or second-generation tissue cysts in the brain. This agreed with Dzierszinski *et al.* (2004), who reported that intra-

cellular bradyzoites are motile within host cells, with the ability to attack surrounding cells and initiate new cysts and that cysts can proliferate by fission, revealing that bradyzoites and cysts are highly dynamic and denoting a mechanism of parasite spreading during chronic infection. Franco et al. (2019) reported that azithromycin treatment was as successful as conventional treatment in human placental villi infected with T. gondii; it considerably reduced T. gondii intracellular proliferation. In the current work, the histopathological examination of mouse brains was used as another evaluation parameter to investigate the efficacy of azithromycin. The histological examination of brain sections of mice from the control positive mice revealed extensive and severe necrosis of neurons and neuronophagia along with the formation of neurofibrillary tangles and numerous large Toxoplasma cysts distributed in the brain parenchyma. There is also major vasculitis and perivasculitis, as well as various localized necrotic necroses along with glial cell infiltration and perivascular cuffing with mononuclear inflammatory cells. This agreed with Waree (2008), who reported that the histopathological hallmarks in the brains of mice with chronic toxoplasmosis were meningeal congestion, with multiple mononuclear cells attacking the meninges and some cuffing of mononuclear cells, around vessels. These changes are the result of malfunction within the blood-brain barrier produced by parasite chemicals that increase the permeability of barrier and enable parasite establishment (Strunk et al, 2014). Astrocytes and glial cells play a dynamic role in the protection against infection by secreting cytokines such as IL-1, IL-6, GM-CSF, IL- 10, IFN-g; and chemotactic cytokines (IP-10 and MCP-1), which prevent lesions and decrease parasite replication (Wilson and Hunter 2004). When the parasite infected a large number of cells, its protective action was reduced or abolished, which aggravates tissue damage and the inflammatory process, favored the creation of tissue cysts. The spiramycin-treated group had the fewest histopathological lesions. The most successful azithromycin regimen was achieved in the 10-day treated mice, which showed mild to severe neuropa thological abnormalities such as neurofibrillary tangles, neuronophagia of degenerated neurons, cerebral blood vessel congestion, and glial cell proliferation. The group treated for only three days had the least effect in terms of severe neuron necrosis, neuronophagia, neurofibrillary tangles, perivascular cuffing with mononuclear cells, meningitis (inflammatory infiltrate in the meninges) associated with the presence of Toxoplasma cysts and different Toxoplasma developmental stages in the cerebral cortex. Azithromycin has been shown to inhibit T. gondii infection and replication in human trophoblastic cells. Franco et al. (2011) reported und that the anti-inflammatory response and enhanced MIF production were important to establish and maintain a healthy in T. gondii infected pregnant mother

Conclusion

Azithromycin treatment for 10 days after the 6^{th} week post-infection in chronic toxoplasmosis proved to be as effective as spiramycin.

Azithromycin reduced the number and size of cysts in brains of infected mice and alleviated the pathological alterations caused by *Toxoplasma gondii* in infected mice.

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Besides, they all equally shared in the theo ritical and practical studies of the paper and approved the final form of the manuscript.

References

Abo-Al-Ela, HG, 2020: Toxoplasmosis and psychiatric and neurological disorders: A step toward understanding parasite pathogenesis. ACS Chem. Neurosci. 11, 16:2393-406

Alday, PH, Doggett, JS, 2017: Drugs in develo pment for toxoplasmosis: Advances, challenges, and current status. Drug Des. Devel. Ther. 11: 273-93

Araujo, FG, Guptill, DR, Remington, JS, 1988: Azithromycin, a macrolide antibiotic with

potent activity against Toxoplasma gondii. Antimicrob. Agents Chemother. 32, 5:755-7.

Araujo, FG, Remington, JS, 1991: Synergistic activity of azithromycin and gamma interferon in murine toxoplasmosis. Antimicrob. Agents Chemother. 35:1672–1673

Araujo, FG, Shepard, RM, Remington, JS, 1991: In vivo activity of the macrolide antibiotics azithr- omycin, roxithromycin and spiramycin against Toxoplasma gondii. Eur. J.Clin. Microbiol. Infect. Dis. 10:519-24

Balaskas, K, Vaudaux, J, Boillat, N, Guex-Crosier, Y, 2012: Azithromycin versus sulfadiazine and pyrimethamine for non-vision-threatening toxoplasmic retinochoroiditis: A pilot study. Med. Sci. Monit. 18, 5:CR296-302.

Beckers, CJ, Roos, DS, Donald, RG, Luft, BJ, Schwab, JC, Cao, Y, *et al*, 1995: Inhibition of cyto-plasmic and organellar protein synthesis in *Toxoplasma gondii*: Implications for the target of macroli-de antibiotics. J. Clin. Invest. 95, 1:367-76

Bosch-Driessen, LH, Verbraak, FD, Suttorp-Schulten, MS, *et al*, 2002: A prospective, randomized trial of pyrimethamine and azithromycin vs pyrimethamine and sulfadiazine for the treatment of ocular toxoplasmosis. Am. J. Ophthalmol. 134:34-40

Chamberland, S, Kirst, HA, Current, WL, 1991: Comparative activity of macrolides against *Toxoplasma gondii* demonstrating utility of an in vitro microassay. Antimicrob. Agents Chemother. 35, 5:903-9

Degerli, K, Kilimcioglu, AA, Kurt, O, Tamay, AT, Ozbilgin, A, 2003: Efficacy of azithromycin in a murine toxoplasmosis model, employing a *Toxoplasma gondii* strain from Turkey. Acta. Trop. 88:45-50

Derouin, F, 1995: New pathogens and mode of action of azithromycin: *Toxoplasma gondii*. Pathol. Biol. 43:561-4

Djurković-Djaković, O, Milenković, V, Nikolić, A, Bobić, B, Grujić, J, 2002: Efficacy of atovaquone combined with clindamycin against murine infection with a cystogenic (Me49) strain of *Toxoplasma gondii*. J. Antimicrob. Chemother. 50, 6:981-7.

Duffy, AR, O'Connell, JR, Pavlovich, M, Ryan, KA, Lowry, CA, et al, 2019: *Toxoplasma gondii* sero-intensity and seropositivity: Heritability and household-related associations in the Old Order Amish. Inter. J. Environ. Res. Publ. Hlth. 16, 19: 3732. 10.3390/ijerph16193732. **Dumas, JL, Chang, R, Mermillod, B, Piguet, PF, Comte, R, Pechère, JC, 1994:** Evaluation of the efficacy of prolonged administration of azithromycin in a murine model of chronic toxo plasmosis. J. Antimicrob. Chemother. 34, 1:111-8

Dunay, IR, Gajurel, K, Dhakal, R, Liesenfeld, O, Montoya, JG, 2018: Treatment of toxoplasmosis: Historical perspective, animal models, and current clinical practice. Clin. Microbiol. Rev. 31, 4:e00057-17

Dzierszinski, F, Nishi, M, Ouko, L, Roos, DS, 2004: Dynamics of *Toxoplasma gondii* differentiation. Eukaryot. Cell 3, 4:992-1003

El-Beshbishi, SN, Elzeky, SM, Atia, RA, Abdalaziz, KF, El-Tantawy, NL, 2020: Toxoplasmosis among Egyptian children with neurological disorders: Developmental and risk factors analysis. PUJ 13, 3:190-6 Online ISSN: 2090-2646.

Feldman, D, Ganon, J, Haffman, R, Simpson, J, 2003: The Solution for Data Analysis and Presentation Graphics. The 2nd Edition, Abacus Lancripts, Inc., Berkeley, USA.

Franco, PS, Gois, PS, de Araújo, TE, da Silva, RJ, de FreitasBarbosa, B, *et al.* 2019: Brazilian st-rains of *Toxoplasma gondii* are controlled by azithromycin and modulate cytokine production inhum-an n placental explants. J. Biomed. Sci. 26, 1: 1-13.

Franco, PS, Gomes, AO, Barbosa, BF, Angeloni MB, Silva NM, *et al*, 2011: Azithromycin and spiramycin induce anti-inflammatory response in human trophoblastic (BeWo) cells infected by *Toxoplasma gondii* but, are able to control infection. Placenta 32(11):838–844.

Gad, WA, Etman, RH, Awad, W, Abdel-Razik, KA, Soror, AH, *et al*, 2022: Immunological, histopathological, molecular identification & genotyping of *Toxoplasma gondii* in small ruminants in Egypt. Pak. J. Biol. Sci. (PJBS) 25, 2: 144-53.

Grujić, J, Djurković-Djaković, O, Nikilić, A, Klun, I, Bobić, B, 2005: Effectiveness of spiramycin in murine models of acute and chronic toxoplasmosis. Int. J. Antimicrob. Agents 25, 3: 226-30.

Harr, T, French, LE, 2010: Toxic epidermal necrolysis and Stevens-Johnson syndrome. Orphanet. J. Rare Dis. Dec 16; 5:39. doi: 10.1186/1750-1172-5-39.

Hernandez, AV, Thota, P, Pellegrino, D, Pasupuleti, V, Benites-Zapata, VA, *et al*, 2017: A systematic review and meta-analysis of the relative efficacy and safety of treatment regimens for HIV-associated cerebral toxoplasmosis: Is trimethoprim-sulfamethoxazole a real option? HIV Med. 18, 2:115-24

Kadappu, KK, Nagaraja, MV, Rao, PV, Shastry, BA, 2002: Azithromycin as treatment for cryptosporidiosis in human immunodeficiency virus disease. J. Postgrad. Med. 48, 3:179-81

Kanoh, S, Rubin, BK, 2010: Mechanisms of action and clinical application of macrolides as immuno-modulatory medications. Clin. Microbiol. Rev. 23, 3:590-615

Lashay A, Mirshahi A, Parandin N, Esfahani, HR, Mazloumi, M, *et al*, 2016: A prospective randomized trial of azithromycin versus trimethoprim/sulfamethoxazole in treatment of toxoplasmic retinochoroiditis. J. Curr. Ophthalmol. 29: 120-5

Lescano SA, Amato Neto V, Chieffi PP, Bezerra, RC, Gakiya, E, *et al*, 2004: Evaluation of the efficacy of azithromycin and pyrimethamine, for treatment of experimental infection of mice with *Toxoplasma gondii*cystogenic strain. Rev. Soc. Bras. Med. Trop. 37, 6:460-2.

Maleki, B, Ahmadi, N, Olfatifar, M, Gorgipour, M, Taghipour, A, Abdoli, A, *et al*, 2021: *Toxoplasma* oocysts in the soil of public places worldwide: A systematic review and meta-analysis. Trans. R. Soc. Trop. Med. Hyg. 115, 5: 471-81

McMullan, BJ, Mostaghim, M, 2015: Prescribing azithromycin. Aust. Prescri. 38, 3: 87-9. Molefe, NI, Musinguz, PS, Kondoh, D, Watanabe, K, et al, 2019: Short- and long-term effects of

orally administered azithromycin on *Trypanosoma brucei brucei* infected mice. Exp. Parasitol. 199:40-6

Montoya, JG, Remington, JS, 2008: Management of *Toxoplasma gondii* infection during pregnancy. Clin. Infect. Dis. 47, 4:554-66

Morsy, TA, Hussein, HE, Morsy, ATA, 2022: TORCH infections, pathogenicity & mortality assessments. JESP 52, 1:53-70.

Srivastava, P, Bhengraj, AR, Jha, HC, Vardhan, H, Jha, R, Singh, LC, *et al*, 2012: Differing effects of azithromycin and doxycycline on cytokines in cells from *Chlamydia trachomatis*-infected women. DNA Cell Biol. 31, 3:392-401

Strunk, T, Inder, T, Wang, X, Burgner, D, Mallard, C, Levy O, 2014: Infection-induced inflammation and cerebral injury in preterm infants. Lancet Infect. Dis. 14, 8:751-62

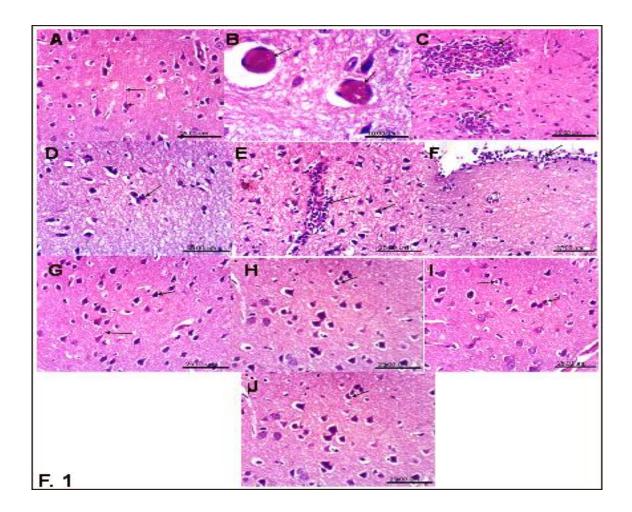
Kamel, AM, Monem, MSA, Sharaf, NA, Magdy, N, Farid, SF, 2022: Efficacy and safety of azithromycin in Covid-19 patients: A systematic review and meta-analysis of randomized clinical trials. Rev. Med. Virol. Jan; 32(1):e2258. doi: 10.1002/rmv.2258.

Waree, CF, 2008: TNF-related cytokines in immunity. In: Paul, W.E. (ed.) Fundamental Immunology. 6th ed. Philadelphia: Lippincott Williams and Wilkins 776-803

Wilson, EH, Hunter, CA, 2004: The role of astrocytes in the immunopathogenesis of toxoplasmic encephalitis. Int. J. Parasitol. 34, 5:543-8

Explanation of figures

Fig. 1: Mice brain to assess azithromycin treating cerebral toxoplasmosis (Me 49 non-virulent strain) lesions (H & E stain). A- Infected nontreated showed necrosis of neurons (short arrow) with appearance of neurofibrillary tangles (long arrow). b-c Infected non-treated showed *Toxoplasma* cysts (arrows) and marked vasculitis and perivasculitis(arrows), respectively. d-f- Azithromycin treated for 3 days showed necrosis of neurons and neuronophagia (arrow), necrosis of neurons (short arrow) and perivascular cuffing with mononuclear cells (long arrow), and meningitis, respectively, with inflammatory infiltrate in meninges. g-i- Azithromycin treated for 10 days showed necrosis of neurons (short arrow) with appearance of neurofibrillary tangles (long arrow), necrosis of neurons (short arrow) and neuronophagia (long arrow), focal gliosis (arrow), respectively. J- Spiramycin treated (after 6thweek PI for 2 weeks) showed degeneration and necrosis of some neurons (arrow) (scale bar 25um).



Explanation of figures

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Group	Drugs	Dose/day	Treatment	Dose	No of <i>T. gondii</i> cyst			size of T. gondii cyst		
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